

Prediction of response to pegylated interferon plus ribavirin in HIV/hepatitis C virus (HCV)-coinfected patients using HCV genotype, *IL28B* variations, and HCV-RNA load

Karin Neukam¹, Angela Camacho², Antonio Caruz³, Norma Rallón⁴, Almudena Torres-Cornejo^{5,6}, Jürgen K. Rockstroh⁷, Juan Macías¹, Antonio Rivero², José M. Benito⁴, Luis F. López-Cortés^{5,6}, Jacob Nattermann⁷, Jesús Gómez-Mateos¹, Vicente Soriano⁴, Juan A. Pineda^{1,*}

¹Unit of Infectious Diseases and Microbiology, Hospital Universitario de Valme, Seville, Spain; ²Unit of Infectious Diseases, Maimonides Institute for Biomedical Research (IMIBIC), Hospital Universitario Reina Sofia, Cordoba, Spain; ³Immunogenetics Unit, Faculty of Sciences, Universidad de Jaen, Jaen, Spain; ⁴Department of Infectious Diseases, Hospital Carlos III, Madrid, Spain;

⁵Unit of Infectious Diseases, Hospitales Universitarios Virgen del Rocío, Seville, Spain;

⁶Instituto de Biomedicina de Sevilla, Seville, Spain; ⁷Department of Medicine I, University of Bonn, Bonn, Germany

Background & Aims: This study aimed at developing a predictive algorithm based on interleukin 28B (*IL28B*) genotype, hepatitis C virus (HCV) genotype, and plasma HCV-RNA load, which could accurately allow us to define the probability of response to pegylated interferon (Peg-IFN) plus ribavirin (RBV) therapy in HIV/HCV-coinfected patients.

Methods: Five hundred and twenty-one treatment-naive HIV-infected patients, who initiated HCV therapy with Peg-IFN/RBV, were analysed in an on-treatment basis. Patients were categorized as unlikely responders, uncertain responders, and anticipated responders (<20%, 20–60%, and >60% probability to achieve SVR, respectively).

Results: HCV genotype, baseline HCV-RNA load, and *IL28B* genotype were confirmed as independent predictors of SVR in a logistic regression analysis. A stepwise algorithm based on these three variables was created based on 321 patients and evaluated in the remaining 200 patients. Unlikely responders included patients with genotype 1 or 4, HCV-RNA load $\geq 600,000$ IU/ml, and rs12979860 non-CC (rate of SVR: 17.3%). Anticipated responders were those with HCV genotype 2–3, patients harboring HCV genotype 4 and *IL28B* CC, as well as those who simultaneously bore HCV genotype 1, HCV-RNA load <600,000 IU/ml, and *IL28B* CC (rate of SVR 74.1%, 77.8%, and 64.4%, respectively). The area under the receiver operating characteristic curve of the model was 0.77 (0.733–0.814).

Conclusions: The combined use of *IL28B* genotype, HCV genotype, and HCV-RNA load enables to easily identify patients with a high and very low likelihood of SVR. HCV therapy could be deferred in the latter patients, until more effective options are available, at least if they do not show advanced liver fibrosis.

© 2011 European Association for the Study of the Liver. Published by Elsevier B.V. All rights reserved.

Introduction

Infection with the hepatitis C virus (HCV) is frequently observed in the HIV-infected population due to shared routes of transmission [1]. The evolution of HCV-related hepatic disease is accelerated in the HIV-infected population, and hepatic complications currently represent a major cause of death among these patients in Western countries [2,3]. Successful anti-HCV therapy is associated with a regression in fibrosis evolution [4], as well as with a lower incidence of complications due to liver-related mortality [3,5]. However, the overall response to the current standard anti-HCV therapy with pegylated interferon (Peg-IFN) plus ribavirin (RBV) is particularly low in HIV-coinfected patients [6–8].

Recent studies have demonstrated that the single nucleotide polymorphism (SNP) rs12979860 near the interleukin 28B (*IL28B*) gene is a strong independent predictor of treatment response in HCV-monoinfected [9–12] and HIV/HCV-coinfected [13,14] patients. The consideration of *IL28B* genotype along with other well-defined pre-treatment determinants of response, such as HCV baseline viral load and HCV genotype, may improve the accuracy of sustained virologic response (SVR) prediction. Indeed, the combination of these three parameters, along with liver stiffness measured by transient elastography, have demonstrated a high diagnostic performance [15]. Thus, a predictive model (Prometheus) [15] has been developed accordingly. However, the Prometheus model does not distinguish between HCV genotype

Keywords: Hepatitis C; Interleukin 28B; Interferon; Ribavirin; Genotype.
Received 8 August 2011; received in revised form 2 November 2011; accepted 17 November 2011; available online 13 December 2011

* Corresponding author. Address: Unidad Clínica de Enfermedades Infecciosas y Microbiología, Hospital Universitario de Valme, Avda. de Bellavista, 41014 Sevilla, Spain.

E-mail address: japineda@telefonica.net (J.A. Pineda).

Abbreviations: HCV, hepatitis C virus; Peg-IFN, pegylated interferon; RBV, ribavirin; SNP, single nucleotide polymorphism; *IL28B*, interleukin 28B; SVR, sustained virologic response; DAA, directly acting agents.



ELSEVIER

1 and 4 and requires fibrosis assessment by transient elastography, a procedure that is not available worldwide.

In a few years, directly acting agents (DAA) against HCV will be commercially available for the treatment of HIV/HCV-coinfected patients. The approval of protease inhibitors, the drug family that will be available earlier [16], is restricted to HCV genotype 1. However, other families with a broader spectrum of activity, such as nucleoside analogue polymerase or NS5A inhibitors, will likely be part of the anti-HCV armamentarium not much later. It is then conceivable that we will be able to use highly active anti-HCV therapies for most HIV/HCV-coinfected patients in a few years. Therefore, in the meantime, the accurate prediction of SVR to Peg-IFN plus RBV will be particularly important. Indeed, it is reasonable to defer therapy until more effective options are available in patients who are unlikely to respond to Peg-IFN plus RBV, at least if they do not show advanced fibrosis. Conversely, patients who have a high likelihood of achieving SVR to Peg-IFN plus RBV may be candidates for immediate treatment. Hence, reliable predictive tools that use a combination of accessible tests, which may accurately foresee SVR in a large proportion of patients, are needed.

The aim of this work was to elaborate an algorithm that could allow to define the probability of achieving SVR to HCV treatment with Peg-IFN plus RBV using viral genotype, *IL28B* genotype, and baseline plasma HCV-RNA without requiring transient elastography in HIV/HCV-coinfected subjects.

Patients and methods

Study population

The study population consisted of HIV/HCV-coinfected patients from four Spanish and one German cohorts prospectively followed in the Infectious Diseases Unit of three university hospitals in Southern Spain, a hospital in Madrid, Spain, and a university hospital in Bonn, Germany, from June 2000 to May 2010. All patients belonging to these cohorts were included in this study if they met the following criteria (i) older than 18 years; (ii) completion of a full course of anti-HCV therapy with Peg-IFN plus RBV; (iii) available data concerning HCV and rs12979860 genotype determinations, as well as a baseline plasma HCV-RNA load quantification. Further details of these populations have been reported elsewhere [13,14,17,18]. A whole blood sample was collected at baseline from all patients and cryopreserved at -80°C for genetic determinations. The programmed visits were scheduled at 4, 12, 24, 36, 48, and 72 weeks (if applicable), as well as 24 weeks after the planned treatment discontinuation date. At each visit, clinical, biochemical, and hematological assessments were carried out.

Drug therapy

All patients received a weekly injection of Peg-IFN α -2a or 2b, at doses of 180 μg or 1.5 $\mu\text{g}/\text{kg}$, respectively. Additionally, 800–1200 mg of oral RBV, according to body weight, was given daily. The treatment duration was 48–72 weeks, with the exception of HCV genotype 2 or 3 carriers who were treated for 24 weeks, if they showed undetectable plasma HCV-RNA load at week 4. Therapy was discontinued in non-responders. Treatment duration and stopping rules were applied according to recommendations of international guidelines in force at the moment of treatment [19,20]. Dose adjustments and the use of granulocyte colony-stimulating factor and erythropoietin were applied according to the decision of the caring physician.

Genetic determinations

The genotype of the *IL28B* SNP rs12979860 was determined as described previously [13,14,18]. In brief, DNA was isolated from whole blood samples and the SNP was genotyped using a custom TaqMan genotyping assay (Applied Biosystems, Foster City, California, USA) or a LightSNiP-Typing Assay (TIB MOLBIOL,

Germany), according to the availability at the corresponding hospital. Likewise, HCV genotyping and determination of plasma HCV viral load were carried out as described elsewhere [13,14,18].

Definition of response

SVR was defined as undetectable HCV-RNA 24 weeks after end of treatment. SVR was assessed in an on-treatment approach, i.e. patients who discontinued treatment voluntarily or due to adverse effects were excluded from analysis.

Data analysis

A descriptive analysis of the data of the study population parameters was conducted. To confirm that HCV viral genotype, rs12979860 genotype, and baseline HCV-RNA load were independently associated with SVR in the study population, binary logistic univariate, and a stepwise multivariate logistic regression analysis were performed.

The patients were randomly split by the statistical software in a 60/40 ratio to obtain two groups, one for the elaboration of the predictive algorithm and the other for its validation. Baseline characteristics of the two groups were compared using the Chi-square test for categorical variables and the Mann-Whitney U-test for continuous variables, respectively. All individuals were categorized in sub-groups, firstly according to HCV genotype, secondly to the baseline HCV-RNA load and finally according to whether they were rs12979860 CC carriers or not. For each group, the rate of SVR was calculated. Patients with a likelihood of SVR of at least 60% were considered *anticipated responders*. Those who had a probability lower than 20% to achieve SVR were classified as *unlikely responders*. The remaining patients were considered *uncertain responders*. Considering SVR as the outcome variable, the predictive capacity of the algorithm was analyzed by means of receiver operator characteristic (ROC) curves generated from the model in the two groups. Anticipated responders developing SVR and unlikely responders who failed to achieve it were considered as patients correctly classified. Anticipated responders without SVR and unlikely responders showing SVR were considered failures of the algorithm. Likewise, the sensitivity, the specificity, the negative predictive value (NPV) and the positive predictive value (PPV) of this algorithm were calculated for those patients that were classified as unlikely or anticipated responders. The statistical analysis was carried out using the SPSS statistical software package release 19.5.0 (IBM Corporation, Somers, NY, USA) and STATA 9.0 (StataCorp LP, College Station, TX, USA).

Ethical aspects

The study was designed and performed according to the Helsinki declaration and was approved by the Ethics Committee of the five participating hospitals. All patients provided written informed consent to participate in this study.

Results

Characteristics of the study population

A total of 521 patients were included in this study. Among them, 236 (45.3%) patients carried rs12979860 genotype CC whereas 285 (54.7%) bore genotype CT or TT. Three hundred and twenty-one patients were randomly selected for the elaboration group, the remaining 200 individuals were included in the validation group. Further baseline characteristics are shown in Table 1.

Response to HCV therapy

In the overall population, 240 (46.1%; CI 95%: 41.7–50.5%) patients showed SVR. SVR was achieved in 151 (63.7%) patients with genotype CC, 69 (30.1%) patients with genotype CT and 21 (37.5%) TT carriers, respectively ($p < 0.0001$). The associations between SVR and HCV genotype, *IL28B* genotype, and baseline plasma HCV-RNA load are shown in Table 2.

Research Article

Table 1. Baseline characteristics of the study population.

Characteristics	Overall population n = 521	Elaboration group n = 321	Validation group n = 200	p
Age (yr)*	42 (39-46)	42 (39-46)	42 (37.8-45)	0.257
Male gender, No. (%)	420 (80.6)	257 (80.1)	163 (80.5)	0.733
IDU [‡] , No. (%)	391 (75)	235 (73.2)	156 (78)	0.252
rs12979860 genotype, No. (%)				
CC	236 (45.3)	146 (45.5)	90 (45)	0.761
TT	56 (10.7)	32 (10)	24 (12)	
CT	229 (44)	143 (44.5)	86 (43)	
HCV genotype, No. (%)				
1	303 (58.2)	183 (57)	120 (60)	0.185
2	7 (1.3)	3 (0.9)	4 (2)	
3	151 (29)	91 (28.3)	60 (30)	
4	60 (11.5)	44 (13.7)	16 (8)	
HCV viral load (log ₁₀ IU/ml)*	6.1 (5.5-6.7)	6 (5.4-6.7)	6.2 (5.5-6.8)	0.383
Undetectable HIV viral load, No. (%) [¶]	324 (77.1)	194 (76.1)	130 (78.8)	0.553
CD4 cell count (cells/ml)*	483 (355-665)	488 (342-700)	475 (370-645)	0.615
ALT (U/L)*	69 (47-111)	69 (47-110)	70 (44-115)	0.872
Advanced liver fibrosis, No. (%) [‡]	171 (39.7)	100 (38)	71 (42.3)	0.420

*Median (Q1-Q3).

[‡]IDU, injection drug user.[¶]Available in 420 patients.[‡]Determined by liver biopsy or transient elastometry, using a cut-off value of 11 kPa, if biopsy was not available; available in 431 patients.*Algorithm for SVR prediction*

To develop the predictive algorithm, the patients of the elaboration group were categorized according to the three predictors of treatment response. The rates of SVR for each resulting category are shown in Fig. 1A. Thus, 132 (41.1%) patients were classified as anticipated responders. This group of patients included three subgroups: the first subgroup including all patients with viral genotype 2 or 3, regardless of the *IL28B* genotype and the baseline HCV-RNA load. The second one including patients with a viral genotype 4, who carried *IL28B* genotype CC, irrespective of the baseline HCV-RNA load, and the third subgroup including patients bearing HCV 1, with *IL28B* genotype CC and who had a baseline HCV-RNA load <600,000 IU/ml. Likewise, 87 (27.1%) individuals were identified as unlikely responders. These patients bore HCV genotypes 1 or 4, presented with a baseline HCV-RNA load of ≥600,000 IU/ml and were *IL28B* genotype CT or TT carriers. The remaining 102 (31.8%) patients were classified as uncertain responders. The predictive values of the algorithm in the elaboration population are shown in Table 3.

In the validation group, 90 (45%) patients showed a response rate of >60% and were classified as likely responders, while 63 (31.5%) were identified as unlikely responders (Fig. 1B). Forty-seven (23.5%) individuals were classified as uncertain responders. The predictive performance of the algorithm in this group was similar to that observed in the elaboration group (Fig. 2 and Table 3).

Frequency of the response categories

The frequency and the rate of SVR of the three categories of response were estimated in the overall population. Thus, 222

(42.6%) patients were classified as anticipated responders to therapy with Peg-IFN and RBV. The patients with viral genotype 2 or 3 showed a SVR rate of 74.1% (CI 95%: 66.5–80.7%), those bearing viral genotype 4 and *IL28B* genotype CC showed an SVR rate of 77.8% (CI 95%: 52.4–93.6%), while in patients harboring HCV 1, *IL28B* genotype CC, and baseline HCV-RNA load <600,000 IU/ml, the SVR rate was 67.4% (CI 95%: 52–80.5%). The overall SVR rate in the group of anticipated responders was 73% (CI 95%: 66.6–78.7%). Sixty (27%) patients in this group did not achieve an SVR and thus were misclassified. One hundred and fifty (28.8%) patients were identified as unlikely responders. Among them, 23 carriers (17.6%; CI 95%: 11.5–25.2%) of genotype 1, and three carriers (15.8%; CI 95%: 3.4–39.6%) of genotype 4 reached an SVR, while 124 (82.7%; CI 95%: 75.6–88.4%) patients did not reach an SVR. The misclassification rate was 17.3%, according to 26 patients showing SVR. A total of 149 (28.6%) patients were classified as uncertain responders. The overall rate of SVR in this subpopulation was 34.9% (CI 95%: 27.3–43.1%).

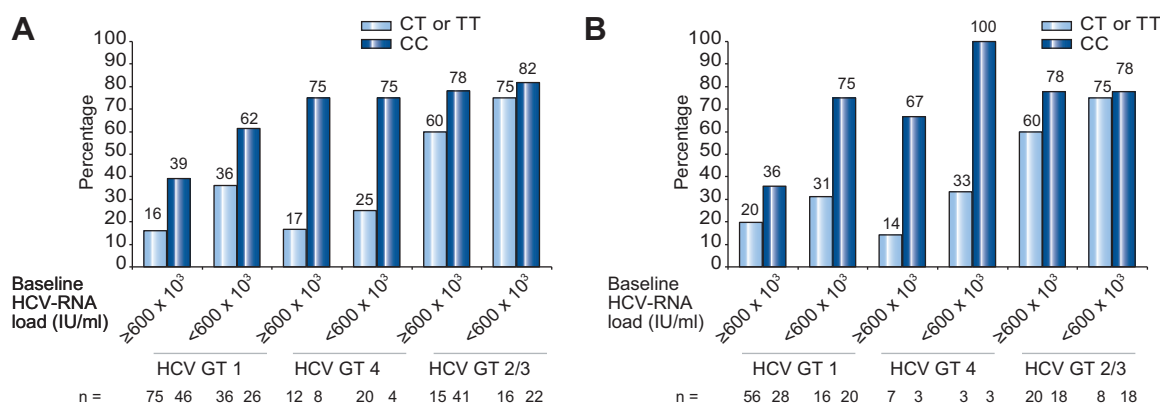
Discussion

The combined use of *IL28B* genotype, HCV genotype, and baseline HCV-RNA load enables to identify patients with a high and a very low likelihood of response to anti-HCV treatment with Peg-IFN and RBV. Using these three variables, we developed an algorithm to predict the rate of SVR which may be applied in a broad range of institutions. This algorithm may help clinicians to make the decision of initiating treatment immediately with Peg-IFN plus RBV or deferring therapy, until more effective options are available.

Table 2. Associations between SVR and HCV genotype, *IL28B* genotype and baseline HCV viral load in the univariate and multivariate analysis (n = 521).

Variable	SVR, No. (%)	<i>p</i> univariate	Adjusted OR (95% CI)*	<i>p</i> multivariate
HCV genotype				
1	100 (33)	<0.0001		<0.0001
2-3	117 (76.6)		4.889 (3.12-7.65)	
4	23 (38.3)			
<i>IL28B</i> genotype				
CC	151 (63.7)	<0.0001	3.312 (2.23-4.9)	<0.0001
TT or TC	89 (31.3)			
Baseline HCV-RNA load (IU/ml)				
<600,000	111 (57.8)	<0.0001	2.192 (1.46-3.29)	<0.0001
≥600,000	129 (39.2)			

*CI, confidence interval.

**Fig. 1. Rates of SVR according to viral genotype (GT), *IL28* genotype, and baseline HCV-RNA load. (A) Elaboration group (n = 321); (B): validation group (n = 200).****Table 3. Predictive performance of the algorithm in patients classified as anticipated or unlikely responders.**

	Overall population (n = 521)	Elaboration group (n = 321)	Validation group (n = 200)
Sensitivity (%)	86.2	87.3	84.6
Specificity (%)	67.4	67	68
Positive predictive value (%)	73	72.7	73.3
Negative predictive value (%)	82.7	83.9	81
Misclassified patients, n (%)	86 (23.1)	50 (22.8)	36 (23.5)

Pre-therapy prediction of SVR is a major challenge in the current clinical practice. The data presented in this study may help to reduce the proportion of treatment failure. Thus, the algorithm allows the identification of patients with a probability of less than 20% to achieve SVR. In the unlikely responders (Fig. 3), it could be considered to defer HCV therapy until more effective options are available, provided that significant fibrosis is absent. Although HIV/HCV-coinfected patients progress considerably fast to higher fibrosis stages, approximately 50% do not show progression of one fibrosis stage in a three-year period [21]. Accordingly, liver fibrosis progression monitoring is an alternative choice, until new drugs can be prescribed to HIV/HCV-coinfected

patients or progression of fibrosis is detected. On the other hand, patients that are classified as anticipated responders (Fig. 3) have an overall probability of 73% to be successfully treated with Peg-IFN/RBV. Therefore, in these patients it would be reasonable to start the bitherapy with Peg-IFN/RBV immediately. In the remaining patients (Fig. 3), individualized decisions should be made. Importantly, the predictive algorithm presented here may be useful for a high proportion of HIV/HCV-coinfected patients, as it allowed us to categorize 372 (71.4%) members of the population as anticipated or unlikely responders.

In this predictive algorithm, the parameters taken into account are HCV genotype, *IL28B* genotype, and baseline

Research Article

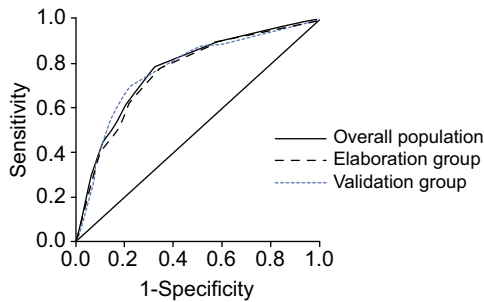


Fig. 2. Diagnostic performance of the predictive index of sustained virological response in accordance with the HCV genotype, the *IL28B* SNP rs12979860, and the baseline viral load. Continuous line: overall population, area under the receiver operating characteristic curve [AUROC (95% CI)]: 0.77 (0.733–0.814), n = 521; dashed line: elaboration group, AUROC (95% CI): 0.77 (0.723–0.826), n = 321; dotted line: validation group, AUROC (95% CI): 0.77 (0.708–0.841), n = 200.

HCV-RNA load (Fig. 3). The predictive performance of this algorithm is somewhat lower than that yielded by the Prometheus model, which, in addition to these three parameters, also evaluates liver stiffness, as determined by transient elastometry [15]. However, the Prometheus index does not differentiate between genotype 1 and 4. This is a limitation, because genotype 4 may account for up to 15% in HIV/HCV-coinfection in some settings [22] and the rate of SVR in patients with HCV genotype 4 is slightly higher. Furthermore, transient elastometry is expensive and not approval worldwide.

A further procedure to predict treatment success is monitoring viral kinetics during therapy with Peg-IFN/RBV. In this context, viral response at week 4 of treatment has high PPV and NPV [23]. Thus, it has been demonstrated that 95% of the patients who present with rapid virological response (RVR), defined as undetectable plasma HCV-RNA viral load at week 4 of therapy, achieve an SVR [23]. Conversely, a decline in plasma HCV-RNA load of <0.6 log units has an NPV of 96% [23]. Furthermore, a

threefold higher treatment success rate was observed in those patients who reached undetectable HCV-RNA at week 12 [24], while a decline of plasma HCV-RNA lower than 2 log units at this time point has an NPV close to 100% [6]. Nevertheless, very few genotype 1 patients show RVR [23] and less than one third of genotype 1 or 4 carriers present with a decline in plasma viral load of <0.6 log₁₀ IU/ml at week 4 of therapy [23]. Therefore, plasma HCV-RNA decline at week 4 allows us to classify only a minority of patients. The assessment of viral kinetics requires exposure to treatment up to 12 weeks. This implies to administer therapy during the period when side effects are more common and severe. Therefore, whenever possible, it is preferable to apply pre-therapy predictive strategies as that presented in this work to spare treatment in those with lower chance to respond.

This study has some limitations. First, this algorithm may be obsolete relatively soon, since triple therapy with DAA/Peg-IFN/RBV will likely be standard-of-care in the near future. However, NS3 protease inhibitors are specific for HCV genotype 1 and bitherapy with Peg-IFN/RBV will still be the treatment strategy in genotype non-1 carriers for several years. Furthermore, drug-drug interactions with antiretroviral treatment may preclude the use of protease inhibitors in a significant proportion of HIV/HCV-coinfected patients [25] and the rate of SVR will probably be lower in the HIV-coinfected population. Thus, pre-therapy prediction of SVR will likely be more important in HIV-coinfected than in mono-infected patients. Moreover, the thresholds selected here to define the response categories are arbitrary. However, response rates of 20% and 60% are reasonable cut-off points to consider patients as good or poor responders. In addition, the proportion of genotype 4 carriers was considerably small, which reflects the HCV genotype distribution in our environment [26–27] and studies with larger sample sizes of this subset are necessary. In fact, an ongoing multicentric study in our area aims to recruit more patients bearing HCV genotype 4 (unpublished data). Furthermore, the HCV genotype and the baseline viral load are well studied predictors of SVR and might have caused a selection bias in the population of this study. However, the genotype

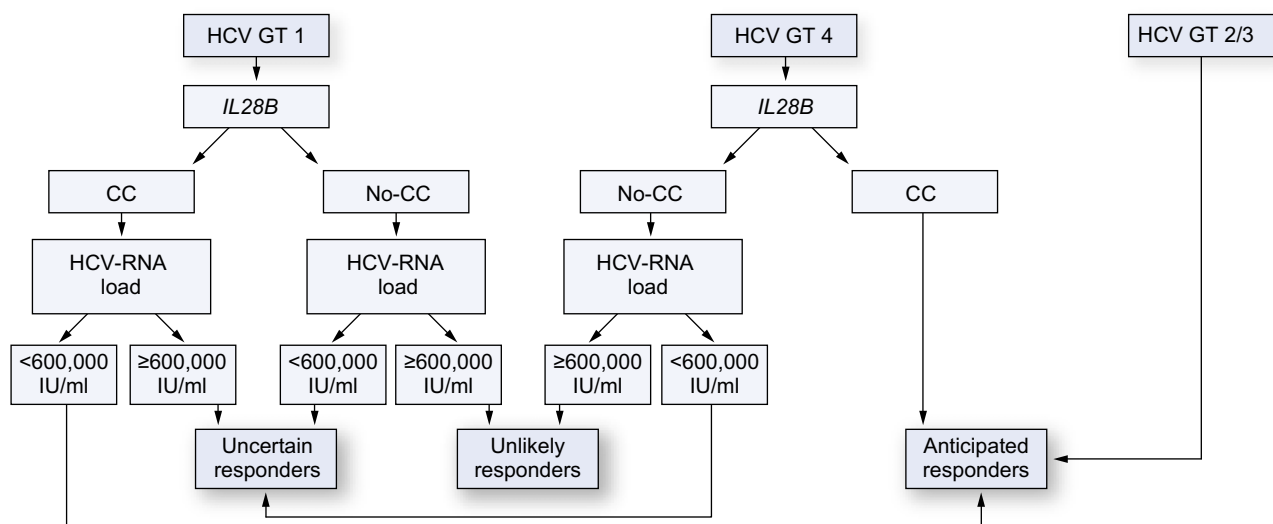


Fig. 3. Stepwise algorithm for the classification of patients as anticipated, unlikely and uncertain responders to bitherapy with peg-IFN/RBV in accordance with the HCV genotype, the baseline HCV load, and the *IL28B* genotype.

distribution reported herein is similar to that reported in epidemiological studies conducted in this environment [26–27]. Therefore, we believe that our results are extrapolable to the global HIV/HCV-coinfected population since no significant deviation can be expected. In any case, the strategy of categorizing subjects according to HCV and *IL28B* genotypes as well as HCV-RNA load allows to define an accurate probability of response in specific subpopulations (Figs. 1 and 2), and enables to use further decision criteria in individual cases, if required. To our knowledge, the sample size of this study is the biggest for a predictive strategy in HIV/HCV-coinfection. Furthermore, this algorithm identifies a proportion of over 70% of patients in whom a categorization as unlikely or anticipated responders can be made, with a fairly low rate of misclassification, without needing fibrosis assessment. Finally, the model was validated by a separate group of patients. These represent the strengths of this study.

In conclusion, this work presents a simple and reliable pre-therapy tool to identify unlikely and anticipated responders to treatment with Peg-IFN plus RBV in HIV/HCV-coinfected patients, including three blood parameters. Two of these parameters were routinely used many years ago and the other has been recently incorporated into clinical practice. This tool may be used to select HIV/HCV-coinfected candidates for immediate and, more importantly, deferred therapy against HCV and it is able to identify as anticipated or unlikely responders in up to approximately three quarters of patients.

Financial support

This study was partly supported by Grants from the Spanish Health Ministry (ISCIII-RETIC RD06/006), the Fundación Progreso y Salud, Consejería de Salud (PI-0247-2010), the Fondo de Investigaciones Sanitarias (PI10/01664 and PI10/01232) and the Fundación para la Investigación y la Prevención del Sida en España (121001/10, 360799/09). K.N. is the recipient of a 'Sara Borrell' postdoctoral perfection Grant from the Instituto de Salud Carlos III (SCO/523/2008). A.R. is the recipient of a research extension Grant from the Fundación Progreso y Salud, Consejería de Salud de la Junta de Andalucía (AI-0011-2010). J.A.P. is the recipient of a research extension Grant from the Fundación Progreso y Salud de the Consejería de Salud de la Junta de Andalucía (AI-0021).

Conflict of interest

The authors who have taken part in this study declared that they do not have anything to disclose regarding funding or conflict of interest with respect to this manuscript.

References

- Alter MJ. Epidemiology of viral hepatitis and HIV co-infection. *J Hepatol* 2006;44:S6–S9.
- Weber R, Sabin CA, Friis-Møller N, Reiss P, El-Sadr WM, Kirk O, et al. Liver-related deaths in persons infected with the human immunodeficiency virus: the D:A:D study. *Arch Intern Med* 2006;166:1632–1641.
- Pineda JA, García-García JA, Aguilar-Guisado M, Rios-Villegas MJ, Ruiz-Morales J, Rivero A, et al. Clinical progression of Hepatitis C Virus-related chronic liver disease in human immunodeficiency virus-infected patients undergoing highly active antiretroviral therapy. *Hepatology* 2007;46:622–630.
- Macías J, del Valle J, Rivero A, Mira JA, Camacho A, Merchante N, et al. Changes in liver stiffness in patients with chronic hepatitis C with and without human immunodeficiency virus coinfection treated with pegylated interferon plus ribavirin. *J Antimicrob Chemother* 2010;65:2204–2211.
- Berenguer J, Alvarez-Pellicer J, Martín PM, López-Aldeguer J, Von-Wichmann MA, Quereda C, et al. Sustained virological response to interferon plus ribavirin reduces liver-related complications and mortality in patients coinfecting with human immunodeficiency virus and hepatitis C virus. *Hepatology* 2009;50:407–413.
- Torriani FJ, Rodriguez-Torres M, Rockstroh JK, Lissen E, González-García J, Lazzarin A, et al. Peginterferon alfa-2a plus ribavirin for chronic hepatitis C virus infection in HIV-infected patients. *N Engl J Med* 2004;351:438–450.
- Pineda JA, Mira JA, Gil IL, Valera-Bestard B, Rivero A, Merino D, et al. Influence of concomitant antiretroviral therapy on the rate of sustained virological response to pegylated interferon plus ribavirin in hepatitis C virus/HIV-coinfected patients. *J Antimicrob Chemother* 2007;60:1347–1354.
- Carrat F, Bani-Sadr F, Pol S, Rosenthal E, Lunel-Fabiani F, Benzekri A, et al. Pegylated interferon alfa-2b vs standard interferon alfa-2b, plus ribavirin, for chronic hepatitis C in HIV-infected patients: a randomized controlled trial. *JAMA* 2004;292:2839–2848.
- Ge D, Fellay J, Thompson AJ, Simon JS, Shianna KV, Urban FJ, et al. Genetic variation in *IL28B* predicts hepatitis C treatment-induced viral clearance. *Nature* 2009;461:399–401.
- Tanaka Y, Nishida N, Sugiyama M, Kurosaki M, Matsuura K, Sakamoto N, et al. Genome-wide association of *IL28B* with response to pegylated interferon-alpha and ribavirin therapy for chronic hepatitis C. *Nat Genet* 2009;41:1105–1109.
- Suppiah V, Moldovan M, Ahlenstiel G, Berg T, Weltman M, Abate ML, et al. *IL28B* is associated with response to chronic hepatitis C interferon-alpha and ribavirin therapy. *Nat Genet* 2009;41:1100–1104.
- Rauch A, Kutalik Z, Descombes P, Cai T, Di Iulio J, Mueller T, et al. Genetic variation in *IL28B* is associated with chronic hepatitis C and treatment failure – A genome-wide association study. *Gastroenterology* 2010;138:1338–1345.
- Pineda JA, Caruz A, Rivero A, Neukam K, Salas I, Camacho A, et al. Variation in interleukin 28B gene predicts response to pegylated interferon plus ribavirin in human immunodeficiency virus/hepatitis C virus-coinfected patients. *Clin Infect Dis* 2010;51:788–795.
- Rallón N, Naggie S, Benito JM, Medrano J, Restrepo C, Goldstein D, et al. Association of a single nucleotide polymorphism near the interleukin-28B gene with response to hepatitis C therapy in HIV/hepatitis C virus co-infected patients. *AIDS* 2010;24:F23–F29.
- Medrano J, Neukam K, Rallón N, Rivero A, Resino S, Naggie S, et al. Modeling the probability of sustained virological response to Therapy with pegylated interferon plus ribavirin in patients coinfecting with hepatitis C virus and HIV. *Clin Infect Dis* 2010;51:1209–1216.
- Sulkowski M, Dieterich D, Sherman K, Rockstroh J, Adda N, Mahnke L, et al. Program and abstracts of the 18th conference on retroviruses and opportunistic infections (Boston). USA: Boston; 2011.
- López-Cortés LF, Valera-Bestard B, Gutiérrez-Valencia A, Ruiz-Valderas R, Jiménez L, Arizcorreta A, et al. Role of pegylated interferon-alpha-2a and ribavirin concentrations in sustained viral response in HCV/HIV-coinfected patients. *Clin Pharmacol Ther* 2008;84:573–580.
- Nischalke HD, Vogel M, Mauss S, Baumgarten A, Lutz T, Danta M, et al. The cytotoxic lymphocyte antigen 4 polymorphisms affect response to hepatitis C virus-specific therapy in HIV(+) patients with acute and chronic hepatitis C virus co-infection. *AIDS* 2010;24:2001–2007.
- European AIDS Clinical Society. European AIDS Clinical Society (EACS) guidelines for the clinical management and treatment of chronic hepatitis B and C coinfection in HIV-infected adults. Available from: http://www.europeanaidscinicalsociety.org/guidelinespdf/3_Chronic_HepatitisB_&_C.pdf [accessed July 2011].
- Soriano V, Puoti M, Sulkowski M, Cargnel A, Benhamou Y, Peters M, et al. Care of patients with hepatitis C and HIV co-infection. *AIDS* 2004;18:1–12.
- Macías J, Berenguer J, Japón MA, Girón JA, Rivero A, López-Cortés LF, et al. Fast fibrosis progression between repeated liver biopsies in patients coinfecting with human immunodeficiency virus/hepatitis C virus. *Hepatology* 2009;50:1056–1063.
- Soriano V, Mocroft A, Rockstroh J, Ledergerber B, Knysz B, Chaplinskas S, et al. EuroSIDA Study Group. Spontaneous viral clearance, viral load, and genotype distribution of hepatitis C virus (HCV) in HIV-infected patients with anti-HCV antibodies in Europe. *J Infect Dis* 2008;198:1337–1344.
- Mira JA, Valera-Bestard B, Arizcorreta-Yarza A, González-Serrano M, Torre-Cisneros J, Santos I, et al. Rapid virological response at week 4 predicts

Research Article

- response to pegylated interferon plus ribavirin among HIV/HCV-coinfected patients. *Antivir Ther* 2007;12:523–529.
- [24] Van den Eynde E, Tiraboschi JM, Tural C, Solà R, Mira JA, Podzamczar D, et al. Ability of treatment week 12 viral response to predict long-term outcome in genotype 1 hepatitis C virus/HIV coinfecting patients. *AIDS* 2010;24: 975–982.
- [25] van Heeswijk R, Vandevorde A, Boogaerts G, Vangeneugden T, de Paepe E, Polo R, et al. Program and abstracts of the 18th conference on retroviruses and opportunistic infections (Boston). USA: Boston; 2011.
- [26] Medrano J, Resino S, Vispo E, Madejón A, Labarga P, Tuma P, et al. Hepatitis C virus (HCV) treatment uptake and changes in the prevalence of HCV genotypes in HIV/HCV-coinfected patients. *J Viral Hepat* 2011;18: 325–330.
- [27] Hüppe D, Zehnter E, Mauss S, Böker K, Lutz T, Racky S, et al. Epidemiology of chronic hepatitis C in Germany – an analysis of 10,326 patients in hepatitis centres and outpatient units. *Z Gastroenterol* 2008;46:34–44.